

# Plant-parasitic nematode communities associated with Moringa tree (*Moringa oleifera* Lam.) in western Niger

A. Haougui<sup>1\*</sup>, A. Basso<sup>1</sup>, I. Mossi Maiga<sup>2</sup>

<sup>1</sup>Institut National de la Recherche Agronomique du Niger <sup>2</sup>University of Tillabéri, Niger

#### Abstract

Moringa (Moringa oleifera Lam) is a crop that is becoming more and more important in Niger, but very little attention has been devoted to its pest problems. Nematode fauna survey was undertaken in three important Moringa producing sites in western Niger. Soil and roots samples were taken from the plant's rhizosphere at 20-30-cm deep. Nematological analysis of these samples revealed the presence of 11 genera of plant-parasitic nematodes among which the most frequent and abundant were *Meloidogyne* spp., *Helicotylenchus* spp. and *Hoplolaimus* spp. The average prominence values of these three nematodes were 236.69, 105.25 and 97.93, respectively. In roots, *Meloidogyne* spp. alone represented up to 90% of the plant-parasitic nematode communities. Its average prominent value was 591.46.

Key words: Moringa oleifera, vegetable crop, plant-parasitic nematodes, Niger.



\* **Corresponding author:** Adamou Haougui, E-mail: <u>ahaougui@yahoo.com</u>

# Introduction

Moringa (Moringa oleifera) plant belongs to the Moringaceae family and originates from the Indian subcontinent (Pandey et al., 2011). In Niger, it is grown as a vegetable crop; its leaves are usually consumed boiled; but sometimes they are eaten raw. Because of its high content in micronutrients (calcium, iron, and vitamins) and proteins (Kimba et al., 2010; Price. 2007), moringa can contribute to the nutritional quality improvement of Niger population. Moringa is grown under irrigation as sole crop or in association with other vegetable crops such as onions, lettuce, peppers, tomatoes or squash. The main producing areas are located in Maradi and Niamey regions. The area devoted to moringa was estimated at 100 000 ha (USAID, 2011). In Sahelian conditions only one insect pest and one pathogenic fungus have been reported in moringa. Abass et al. (2007) reported damages caused by insect pests that destroy foliage during the dried season. The main pest is a moth (Noorda blitealis, Lepidoptera, Crambidae) identified for the first time in Niger by Ratnadas et al. (2011). Haougui et al. (2014) identified for the first time **Botriodiplodia** of theobromae responsible the as moringa decline observed in the western part of the country. To date, no information is available on plant parasitic nematodes of Moringa in Niger. Information reported by Prot (1984) showed that this crop, like papaya and baobab tree, can also serve as a reservoir of root-knot nematodes belonging to the genus Meloidogyne. This author noticed that moringa did not show any symptoms

of nematode attacks apart from having root galls. The objective of this study was to identify the plant- parasitic nematodes associated with Moringa in Niamey, one of the two largest producing areas of moringa in Niger.

# Materials and methods

Site characteristics and sampling: The three main producing sites of Moringa in the Western region of Niger, Toulware, Karey-Gorou and Sarando were selected for study (Table 1). In these sites, Moringa coexists with other fruit trees such as mango (Mangifera indica), guajava (Psidium guajava, citrus (Citrus spp.) and hedge trees comprising (Acacia Senegal, Bauhinia rufescens or Prosopis juliflora. In the moringa plots, other vegetable crops such as tomatoes, peppers, eggplant and cucurbits (melon and zucchini) were planted. Ten to fifteen farms were surveyed based on the importance of the sites. Sampling was done using the zig-zag method of Barker (1985). On each plot, samples of soil and roots were taken in the rhizosphere of plants at a depth of 20-30 cm, with a trowel. Each sample (2 kg of wet soil + roots), composed of 5 sub-samples was put in a plastic bag, labeled and then placed in a strong box and transported to the laboratory for nematological analysis.

Table 1: location of study sites.				
Sites	Geographic coordinates			
Toulware	13° 28''44.13''N ; 2° 01' 01.78''E			
Karey-gorou	13 ° 32' 15.91''N ; 2° 00' 23.70''E			
Sarando	13° 35' 14.43'' N ; 1° 55' 49.32''E			

Nematode extraction, identification and data analysis: Nematodes were extracted from the soil and roots using Seinhorst methods (1950 and 1962). The importance of each genus or species of nematode was determined by calculating the mean density of nematodes per site, the relative density, the frequency of occurrence in the samples and the prominent value. The mean density is the average of individuals per  $250 \text{ cm}^3$  of soil or 5 g of root. The relative density of each nematode was calculated using the following formula:

$$RelativeDensity = \frac{Number of nematodes}{Total number of nematodes} x100$$

The frequency (f) is the percentage of samples which contain the given nematode as calculated by the following formula:

$$Frequency = \frac{Number of samples containing a nematode}{Total number of samples} x100$$

The prominent value is given by the formula of De Waele et al. (1998) as follow:

Prominent value = Density x 
$$\frac{\sqrt{\text{(frequency of appearance)}}}{10}$$

# Results

Importance of parasitic nematodes in the soil: The nematological analysis of soil samples showed 11 genera of plantparasitic nematodes in the rhizosphere of moringa. *Meloidogyne* and *Pratylenchus* are respectively sedentary endoparasite and migratory endoparasites, while the other nematodes are ectoparasites. although Scutellonema clathricaudatum has endoparasitic phase. Karey-gorou showed the greatest biological diversity with 10 genera, followed by Sarando and Toulware infested by 9 and 8 kinds of plant-parasitic nematodes, respectively. Seven genera were common to all the three sites. These are Meloidogyne, Xiphinema, Helicotylenchus, Pratylenchus, Hoplolaimus, *Tylenchorhynchus* and Criconemella (Table 2). Analysis of plant-parasitic

nematodes communities of the 3 sites showed that Scutellonema exhibited the highest prominent value (PV) at Toulware followed by Meloidoyne, Hoplolaimus and Helicotylenchus. The PVs were respectively 136.99, 117.06, 105.25 and 93.56. On the other two sites, Meloidogyne recorded the largest PV (204.33 and 388.67), followed by Helicotylenchus (117.79 and 104.48) and Hoplolaimus (115.89 and 72.65). The other nematodes were not abundant in all the 3 sites (Table 3). The figure 1 shows relative densities different the of nematodes in the soil. It appears that the three genera with the highest PVs alone represent up to 65% of the parasitic nematodes at Toulware. Meloidogyne alone accounted for almost a quarter of the nematode population in this site. Hoplolaimus, Meloidovne and Helicotylenchus represented more than 90% of the nematodes at Karey-gorou and Sarando. *Meloidogyne* alone accounted respectively for 39 and 56% of the nematode communities. The most poorly represented genera were

*Tylencorhynchus* (PV = 0.6) in the first site and *Psylenchus* (PV = 0.3) and *Tylenchus* (PV = 0.50) respectively in the second and third sites.

Table 2: distribution of parasitic nematodes by site.

Parasitcnematodesspecies	Toulware	Karey-gorou	Sarando
Meloidogynespp	+	+	+
Xiphinemaelongatum	+	+	+
Helicotylenchusdihystera	+	+	+
Pratylenchusbrachyurus	+	+	+
Hoplolaimuspararobustus	+	+	+
Tylenchorhynchusindicus	+	+	+
Criconemellacurvata	+	+	+
Scutellonemaclathricaudatum	+	-	-
Paratrichodorusminor	-	+	-
Aphelenchussp	-	+	+
Tylenchus sp	-	+	+

#### Table 3: Importance of plant-parasitic nematodes from soil samples of moringa.

Sites	Nematodegenera	Nematode density (Nbr individuals/250 cm <sup>3</sup> )	Frequency (%)	Prominence value
Toulware	Meloidogyne	117.06	100.0	117.06
	Xiphinema	7.44	51.6	5.34
	Helicotylenchus	93.56	100.0	93.56
	Pratylenchus	2.75	72.8	2.35
	Hoplolaimus	105.25	100.0	105.25
	Tylencorhynchus	0.75	64.6	0.60
	Criconemella	1.9375	55.1	1.44
	Scutellonema	152.97	80.2	136.99
	Meloidogyne	204.33	100.0	204.33
	Xiphinema	9.38	17.4	3.92
	Helicotylenchus	134.75	76.4	117.79
	Pratylenchus	16.38	50.2	11.60
Karey-Gorou	Hoplolaimus	133.38	75.5	115.89
	Tylencorhynchus	7.19	56.8	5.42
	Criconemella	2.06	25.0	1.03
	Paratrichodorus	7	61.0	5.47
	Psylenchus	0.56	32.2	0.32
	Scutellonema	8.06	100.0	8.06
Sarando	Meloidogyne	388.67	100.0	388.67
	Xiphinema	11.72	15.7	4.64
	Helicotylenchus	133.23	61.5	104.48
	Pratylenchus	20.47	53.7	15.00
	Hoplolaimus	122.58	35.2	72.67
	Tylencorhynchus	8.98	32.1	5.09
	Criconemella	2.58	52.4	1.87
	Aphelenchus	8.75	21.7	4.07
	Tylenchus	0.7	51.7	0.50

**Importance of parasitic nematodes in the roots:** Only three parasiticnematodes were extracted from the roots of moringa. These were *Meloidogyne*, **Pratylenchus** Scutellonema. and At Sarando, Meloidogyne only and Pratylenvhus were found. Table 4 presents importance of these the

parasitic-nematodes in the roots in the three sites. It appears that *Meloidogyne* was the most important parasitic nematodes in the roots. Its prominent values were 355.00, 908.22 and 511.15 at Toulware, Karey-gorou and Sarando, respectively. *Pratylenchus* and *Scutellonema* were very scarce with relatively very low densities ranging between 8 and 61 individuals / 5 g roots. Figure 2 shows the relative densities of the different parasitic-nematodes found in the roots. *Meloidogyne* species represented most of the endophytic nematode communities as it accounted for 84 to 96% of the total.



Figure 1: Relative densities of plant-parasitic nematodes in moringa rhizosphere.

Sites	Nematodegenera	Nematode density (Nbr individuals/ 5gr of roots <sup>3</sup> )	Frequency (%)	Prominence values
	Meloidogyne	355	100	355.00
Toulware	Pratylenchus	61	55.67	45.51
	Scutellonema	8	23.46	3.87
	Meloidogyne	908	100	908.22
Karey-gorou	Pratylenchus	42	53.12	30.61
, 6	Scutellonema	30	48.05	20.97
Sarando	Meloidogyne	511	100	511.15
	Pratylenchus	22	76.54	19.03

Table 4: Importance of plant-parasitic nematodes from root samples of moringa.



Figure 2: relative densities of plant-parasitic nematodes in moringa plant roots

### Discussion

The results described in this study showed that on all the three sites, 11 genera of plant-parasitic nematodes were associated with moringa. In the same region, 8 to 12 genera of phytonematodes were found on Solanaceous crops and hedge tree species (*Acacia senegal*, *Prosopis juliflora* and *Bauhinia rufescens*), respectively (Haougui et al., 2013a and 2013b). All these nematodes were already reported on vegetable crops by Haougui (1999). The root-knot nematodes, *Meloidogyne* spp., the most devastating group on vegetable crops were found in all sites with high densities. In roots, they constitute the dominant group of parasitic-nematodes communities. In Niger, three species of Meloidogyne, M. incognita (Silva, 2005; Sikora et al., 1988), M. javanica (Haougui, 1999) and M. enterolobii (Nourh, 2012) are known to occur. Haougui et al. (2008) reported yield losses due to M. javanica up to 60% on tomato. They noted that heavy infested soil can lead to total yield loss. However, during our field survey, no symptoms of nematode infection were perceptible on moringa plants, despite the high densities of Meloidoyne in the roots. This is consistent with results reported by Prot (1984) who thinks that moring is only a reservoir of Meloidogyne spp. So, like the hedges species, it contributes to contaminate vegetable crops that are in its immediate vicinity (Senego & Cadet, 2007). This study also confirmed the polyphagous status of *Meloidogyne* spp. that can infect more than 2000 plant species worldwide (Nourh, 2012). In addition, root-knot nematodes have been found in all vegetable producing areas of Niger where they constitute the main limiting factor to crop production (Haougui et al., 2013c and 2013d), especially since they most often act in synergy with some of the vascular disease pathogens. Haougui and Bizo (2009)reported the simultaneous presence of Meloidogyne spp, Ralstonia solanacearum and Fusarium spp on pepper in the Aguie area and they attributed the rapid decline of the crop production in the region of Maradi (Niger) to the action of the parasite complex. Other parasitic nematodes with large prominent values (Helicotylenchus and Hoplolaimus on all three sites and Scutellonema at Toulware), are not very dangerous for vegetable crops despite

their high densities. Some authors considered Helicotylenchus dihysteraas a minor pest and sometimes as an indicator species of mature fallow in which nematode biodiversity is very high (Villenave & Cadet, 1998). In such an environment, the free living nematodes and parasitic nematodes densities are very high (Serigne et al., 2003). Cadet (1998) reported that H. dihystera expressed a moderating effect on the pathogenesis of the communities to which it belongs. According to Rodriguez-Kabana (1987), when some plant-parasitic nematodes species are in a multispecies community, they have no significant depressive effect on the plants' growth. Hoplolaimus is mostly a parasitic nematode observed on millet, sorghum and cowpea in the Sahel area (Baujard et al., 1995). Its occurrence on our sites may be explained by the cultivation of these crops in association with moringa. Scutellonema *clathricaudatum*, the main species of the identified in Niger, causes genus significant damage only to peanuts under rainfed conditions (Baujard & Martiny, 1995; Sharma et al., 1992; Sharma et al., 1990). However, this crop is not grown in vegetable sites even in Maradi, one of the largest peanut basins of Niger. But if one day these lands should be allocated to the production of this crop, special attention should be paid to S. clathricaudatum. The study revealed that the rhizosphere of moringa has а multispecies community of phytopathogenic nematodes, including Meloidogyne spp which is a limiting factor in the vegetable crops production in Niger. So, any development program of moringa should consider its nematotological status in order to avoid associations with vegetable crops. The pathogeny of *Scutellonema clathricaudatum* and *Hoplolaimus pararobustus* on moringa should be tested both in a greenhouse and fields to understand their real effect on this plant.

### Acknowledgement

The authors thank the Director General of Niger Agricultural Research Institute (INRAN) for funding this work. Thanks are also addressed to the moringa producers of Toulware, Karey-gorou and Sarando for all the information they provided.

## References

- Abass TA, Gouzayé A, Woltering L, Pasternak D, 2007. The role of indigenous leafy vegetables on daily diet and rural and urban economy of Niger. Acta Horticulturae **752**: 35–40.
- Baujard P, Martiny B, 1995. Ecology and pathogenicity of the Hoplolaimidae (Nemata) from the sahelian zone of WestAfrica. 3: Scutellonema clathricaudatum Whitehead, 1959. Fundamental and Applied Nematology 18 (4): 347–353.
- Baujard P, Bour E, Martiny B, 1995. Incidence des nématodes phytoparasitaires sur la culture du sorgho dans la zone sahélienne du Sénégal, Afrique de l'Ouest. Afro-Asian Journal of Nematology **5**: 1–10.
- Barker KR, 1985. Sampling nematode communities. In: Barker, KR., Carter KRC, Sasser JN, eds. An advanced

treatise on *Meloidogyne*, Vol. 2: Methodology.North Carolina State University Graphics, Raleigh, North Carolina, USA.

- Cadet P, 1998. Gestion écologique des nématodes phytoparasites tropicaux. Cahiers d'Agriculture **7:** 187-194.
- Cadet P, Sanogo D, 2007. Rôle potentiel des haies vives pour la gestion des nématodes sur les bassins versants de la zone soudano- Sahélienne au Sénégal. Tropicultura **3**; 153–160.
- De-Waele D, McDonald AH, Jordan EM, Orion D, Van Den Berg E, Loots GC, 1998. Plant-parasitic nematodes associated with maize and peal millet in Namibia. African Plant Protection **4**: 113–117.
- Haougui A, 1999. Parasitic nematodes of vegetable crops in Niger: importance and control method by using plants. Dschang, Cameroon, University of Dschang, PhD thesis.
- Haougui A, Sarr E, Alzouma I, 2008. Effet des feuilles sèches de neem (Azadirachta indica A. Juss) et du ricin (Ricinus communis L.) sur le nématode à galles Meloidogyne javanica, parasite de au Niger. Sciences la tomate et Sciences technique, naturelles et agronomie **30 (2)** : 27–35.
- Haougui, A,BizoN, 2009. Characterization of plant parasitic nematode communities associated with of pepper on the Aguie KKM project sites of Maradi, the KKM Project activity report.
- Haougui A, Basso A, Abdou HO, Sidikou RSD, Adam T, 2013a. Characterization of plant-parasitic nematode communities associated with tomato, eggplant and pepper in the suburban area of Niamey

(Niger). International Journal of Agriculture and Crops Science **5(20)**: 2488–2494.

- Haougui A, Sarr E, Balde H, Doumma A, Adam T, 2013b. Characterization of Plant-Parasitic-Nematode Communities Associated with tree Species used in Hedges in Western Niger and their Susceptibility to *Meloidogyne javanica*. Asian Journal of Agriculture and Rural Development **3(6)**: 378–386.
- Haougui A, Kassoum MN, Basso A, Doumma A, Adam T (2013c). Plant parasitic nematode communities associated with pepper crops in Aguie region (Niger Republic). Botany Research International **6** (**1**), 01–06.
- Haougui A, Sarr E, Djibey R (2013d).Biodiversity of plant-parasitic nematodes associated with pepper in the regions of Diffa and Dosso (Niger republic). International Journal of Farming and Allied Sciences **2(15)**: 482– 487.
- Haougui A, Basso A, Mbaye N, Abdoulaye AK, Delmas P, 2014. Alerte: une nouvelle maladie sur le moringa au Niger. Reca-Niger online <u>http://www.recaniger.org/IMG/pdf/REC</u> <u>A moringa diplodia decembre 2014.pdf</u>
- Kimba A, Dan-Marafa B, Abdou H, 2010. Le moringa, une demande forte, une offre insuffisante. Note d'information/filière moringa N°1, RECA, Niamey, Niger.
- Nourh Y, 2012. Prevalence and characterization of plant-parasitic nematodes on eggplant, pepper, tomato and guava in the western part of Niger. Gent, Belgium, University of Gent, Msc. Thesis.

- Pandey A, Pradheep K, Gupta R, Nayar ER, Bhandari DC, 2011. Drumstick tree (*Moringa oleifera* Lam.): a multipurpose potential species in India. Genetic Resources and Crop Evolution **58** (**3**): 453–460.
- Price LM (2007). Le Moringa. ECHO (Ed.), North Fort Myer, USA.
- Prot JC, 1984. Les nématodes phytoparasites des cultures maraîchères. ORSTOM/USAID (Eds.), Dakar, Senegal.
- USAID, 2011. Reducing the risk of acute malnutrition and household food insecurity in Niger through moringa cultivation. Technical repport, February, 2011.
- Ratnadass A, Zakari MO, SalhaH, Minet J, Seyfoulaye A, 2011. Noorda blitealis Walker, un ravageur majeur du moringa au Niger (Lepidoptera, Crambidae). Bulletin de la Société d' Entomologie 116(4): 401–404.
- Rodriguez-Kabana R, Robertson DG, King PS, 1987.Comparison of methyl bromide and other nematicides for control of nematodes in peanut. Annual and Applied Nematology 1: 56–58.
- Seinhorst JW, 1950. The significance of soil conditions on the action of the stem nematode (*Ditylenchus dipsaci*(Kühn) Filipjev). Tijdschrift Plantenziekten **56**: 291–349.
- Seinhorst JW, 1962. Modification of the elutriation method for extracting nematodes from soil. Nematologica 8: 117–128.

- Sharma SB, Subrahmanyam P, Sarr E, 1990. Plant parasitic nematodes associated with grounudnut in Niger. Tropical Pest Management **36**, 71–72.
- Sharma SB, Waliyar F, Subrahmanyam P, Ndunguru BJ, 1992. Role of *Scutellonema clathricaudatum* in etiology of groundnut growth variability in Niger. Plant and Soil **143**: 133–139.
- Sikora RA, Reckhaus P, Adamou E, 1988. Presence, distribution and importance of plant parasitic nematodes in irrigated agricultural crops in Niger. Medical Faculty Landbouwwetenschappen Rijksuniversiteit Gent **53/2b**: 821–834.
- Silva GD, 2005. Contribution à l'étude de *Meloidogyne* et de la nématofaune associée en culture Maraîcher au Niger. Niamey, Niger, Centre Régional AGRHYMET, Msc thesis.
- Serigne TK, Callistus KPO, Albrecht A, 2003.Crop damage by nematodes in improved-fallow fields in western Kenya. Agroforestry Systems **57**: 51–57.
- Villenave C, Cadet P, 1998. Interaction of *Helicotylenchus dihystera*, *Pratylenchus pseudopratensis* and *Tylenchorhynchus gladiolatus* on two plants from the soudano-sahelian zone of zest Africa. Nematologica 2: 31–39.